

Research Article

# Diagnostic accuracy of salivary biomarkers in Alzheimer's disease: A systematic review

Amir A. Majeed<sup>1</sup>, Auday M. Al-Anee<sup>2\*</sup>, Natheer H. Al-Rawi<sup>3</sup>

<sup>3</sup>

1 Department of Oral Diagnostic Sciences, College of Dentistry, University of Baghdad, Iraq.

2 Department of Oral and Maxillofacial Surgery, College of Dentistry, University of Baghdad, Iraq.

3 Department of Oral and Craniofacial Health Sciences, University of Sharjah, UAE.

\*Correspondence: [audayalanee@gmail.com](mailto:audayalanee@gmail.com)

Received date: 01-07-2024

Accepted date: 02-09-2024

Published date: 15-12-2025



Copyright: © 2025 by the authors. Submitted for possible open access publication under the terms and conditions of the Creative Commons Attribution (CC BY) license

(<https://creativecommons.org/licenses/by/4.0/>).

Article DOI



**Abstract:** Background: Alzheimer's disease causes cognitive decline and neuronal death due to amyloid-beta plaques and neurofibrillary tangles. Current diagnostic methods are intrusive, expensive, and few, but early diagnosis improves patient outcomes. Due to its oral-brain axis relationship to the central nervous system, saliva may be a viable source of non-invasive Alzheimer's biomarkers. This systematic review aimed to assess the diagnostic potential of salivary biomarkers for Alzheimer's disease by carefully reviewing the existing data. Methodology: Studies published between January 1, 2008, and September 30, 2023, were the subject of a thorough literature search in the PubMed, Scopus, and Web of Science databases. Relevant data were extracted after studies were evaluated according to predetermined inclusion and exclusion criteria. The Oxford Centre for Evidence-Based Medicine criteria were implemented to evaluate the quality of the studies that were incorporated. Results: A total of 36 studies were included in the systematic review, as they met the inclusion criteria. Alzheimer's disease patients exhibited significantly elevated salivary levels of beta-amyloid-42 and p-tau in comparison to healthy controls. Patients with Alzheimer's disease exhibited non-significant trends in salivary t-tau and lactoferrin. Conclusion: The levels of beta-amyloid-42 and p-tau in the saliva of patients with Alzheimer's disease are significantly altered. Beta-amyloid-42 regard as a best marker in early diagnosis by salivary sample. These findings emphasize the prospect of saliva as a non-invasive source of biomarkers for the diagnosis of Alzheimer's disease. Nevertheless, salivary tests must undergo additional validation in larger, well-characterized cohorts and standardization of methodologies before they can be implemented in clinical practice.

**Keywords:** Alzheimer's disease, Salivary biomarkers, Systematic review, saliva proteomics.

## Introduction

Alzheimer's disease (AD) is a severe neurological disease that gradually deprives persons of their memories, cognitive functions, and finally, their identity. Globally, Alzheimer's disease (AD) is the primary underlying factor contributing to the development of dementia. Hence, Alzheimer's disease imposes a significant burden not only on the individuals affected by it, but also on their family members and the

broader community<sup>(1,2)</sup>. Despite ongoing efforts utilizing advanced technologies, existing diagnostic procedures are unable to detect the early stages of the disease, hence reducing the effectiveness of treatment. Therefore, the primary objective that clinicians should undertake is to promptly diagnose the disease<sup>(3)</sup>.

AD is caused by a complicated and multifaceted pathophysiology that involves the buildup of beta-amyloid (A $\beta$ ) plaques, the presence of phosphorylated tau (p tau) in the brain, damage to synapses, loss of neurons, oxidative stress, neuroinflammation, and impaired metabolism of mitochondria<sup>(4-6)</sup>. The accumulation of degenerative processes results in brain atrophy and the manifestation of dementia symptoms, including memory loss, apathy, and impaired ability to carry out everyday activities<sup>(7)</sup>. The diagnosis of Alzheimer's disease (AD) should rely on the assessment of molecular indicators and clinical symptoms. Standard diagnostic techniques, such as cerebrospinal fluid biomarkers, together with imaging procedures like magnetic resonance imaging (MRI) and positron emission tomography (PET), are used to assess regions of brain shrinkage, impaired metabolism, and aberrant buildup of chemicals<sup>(8-10)</sup>. However, these procedures are invasive and expensive, and a significant number of individuals are unable to utilize them, prompting the exploration of alternative approaches. Several theories have been proposed to explain the emergence of salivary biomarkers related to AD. One theory suggests that neurons may leak biomarkers into the salivary glands because they are near the central nervous system<sup>(11)</sup>. Saliva, being a readily accessible and non-invasive bodily fluid, has demonstrated potential in indicating a range of pathophysiological states, such as gastrointestinal, thyroid, oncological, autoimmune, cardiovascular diseases, and neurological disorders. The oral-brain axis, which involves many routes linking the oral cavity and the central nervous system, presents an opportunity to identify signs of Alzheimer's disease in saliva<sup>(12-16)</sup>.

The primary question that governs this systematic review is: "Is it feasible to employ salivary biomarkers for the diagnosis of Alzheimer's disease with a high degree of reliability?" This review explores the current state of reported evidence to provide a comprehensive report on the most promising salivary biomarkers for AD, their diagnostic capabilities, and the challenges and opportunities that practitioners encounter in this rapidly evolving field. Finally, we aim to expand the boundaries of technology by promoting non-invasive and accessible diagnostic methods to aid in early diagnosis. This, in turn, can be beneficial for treatment and alleviate the significant burden of this disease on the individual, family, and society.

### Search Strategy and Data Extraction

The Preferred Reporting Items for Systematic Reviews and Meta-Analyses (PRISMA) provide a framework to conduct these systematic reviews<sup>(17)</sup>. We conducted a search in the databases of PubMed, Scopus, and Web of Science, specifically looking at the references published between 01/01/2008 and 30/09/2023. In addition, we have executed search strategies. The search terms used were meticulously designed to capture pertinent material.

The Boolean operators utilize the following combinations:

- For PubMed: (saliva) AND (marker OR biomarker OR enzyme OR metabolite OR hormone) AND (Alzheimer).
- For Scopus: (saliva) AND (marker OR biomarker OR enzyme OR metabolite OR hormone) AND (Alzheimer).

- For Web of Science: (saliva) AND (marker OR biomarker OR enzyme OR metabolite OR hormone) AND (Alzheimer).

The search results were then manually refined to exclusively encompass journal articles published on or after January 1st, 2008. It is crucial to emphasize that the search method is organized to encompass topics related to AD. This review being reviewed by two authors and in case of any discrepancies or disagreement the third author would solve these discrepancies. During the systematic screening procedure, two distinct reviewers conducted searches for pertinent publications based on their titles, abstracts, and full texts. Only studies that met the pre-established criteria based on the PICO framework, which stands for Population, Intervention, Comparison and Outcomes were included in this study.

## Materials and Methods

### Inclusion and exclusion criteria

**Population:** Ensured that both genders were represented, and the participants were from all age categories, in this instance, the decision is to preserve the statistical robustness of a randomized controlled trial (RCT) by enrolling a minimum of 15 patients, and preferably more. The studies that had a sample size of less than 15 patients or controls were excluded.

**Intervention:** The primary objective of the systematic review was to address Alzheimer's disease. Excluded studies that examined other diseases, such as Mild Cognitive Impairment (MCI), in order to preserve the specificity of our research inquiry.

**Comparison:** The presence of a non-demented control group was a critical component of the inclusion criteria for the purpose of comparison. Research that did not include a non-demented control group was considered ineligible for inclusion.

**Results:** The primary focus of our review was the alterations in the levels of salivary markers. Excluded studies that examined changes in markers from other biological samples, such as serum, or that investigated the microbiota, as they were not within the scope of the research question.

**Study design:** Incorporated case-control, cohort, and cross-sectional studies to guarantee a comprehensive and robust analysis. Nevertheless, literature reviews, case reports, expert opinions, letters to the editor, and conference reports were excluded.

Furthermore, to guarantee that the most pertinent and recent literature was incorporated, the search was limited to studies published after January 1, 2008. Additionally, this systematic review exclusively analyzed studies that were published in the English language.

This systematic review endeavored to maintain a high level of scientific integrity and relevance by conducting a comprehensive and rigorous analysis of the changes in salivary markers for the diagnosis of Alzheimer's disease in accordance with these well-defined inclusion and exclusion criteria.

**Assessment of the quality of study:** This systematic review utilizes a rigorous evaluation of the quality and reliability of evidence from the studies included. It employs the evidence classification scheme developed by the Oxford Centre for Evidence-Based Medicine (OCEBM)<sup>(18)</sup> which is widely recognized and

respected in the field. The purpose of this five-level scale is to evaluate the accuracy of diagnostic investigations and determine their reliability. Higher levels on the scale indicate stronger and more dependable evidence.

#### Quality Assessment and Critical Appraisal for the Systematic Review of Included Studies

The quality of studies involved in the systematic review was given a score of 0 to 9 using the Newcastle-Ottawa Scale (NOS) <sup>(19)</sup>. The final score was converted into categories of good (7 to 9), fair (4 to 6), and poor (0 to 3) according to the Agency for Healthcare Research and Quality standards (AHRQ).

## Results

### Literature search

Identification Phase: A total of 1,307 records were obtained from the initial search across three databases (PubMed, Scopus, and Web of Science). PubMed recorded 346 records, Scopus recorded 521 records, and Web of Science recorded 440 records. After the removal of duplicate records, there were 549 unique records.

Screening Phase: The titles and abstracts of the 758 remaining records were considered during the screening process. Following this screening process, 616 records were eliminated leaving 142 reports that were submitted for retrieval.

Eligibility Phase: One report out of the 142 report was unable to be retrieved. Consequently, 141 reports were evaluated for eligibility according to the predetermined inclusion and exclusion criteria.

During this audit, a total of 105 reports were eliminated due to the following reasons:

A total of 63 reports were deemed irrelevant to the review.

There were 36 reports specifically related to Parkinson's disease were excluded.

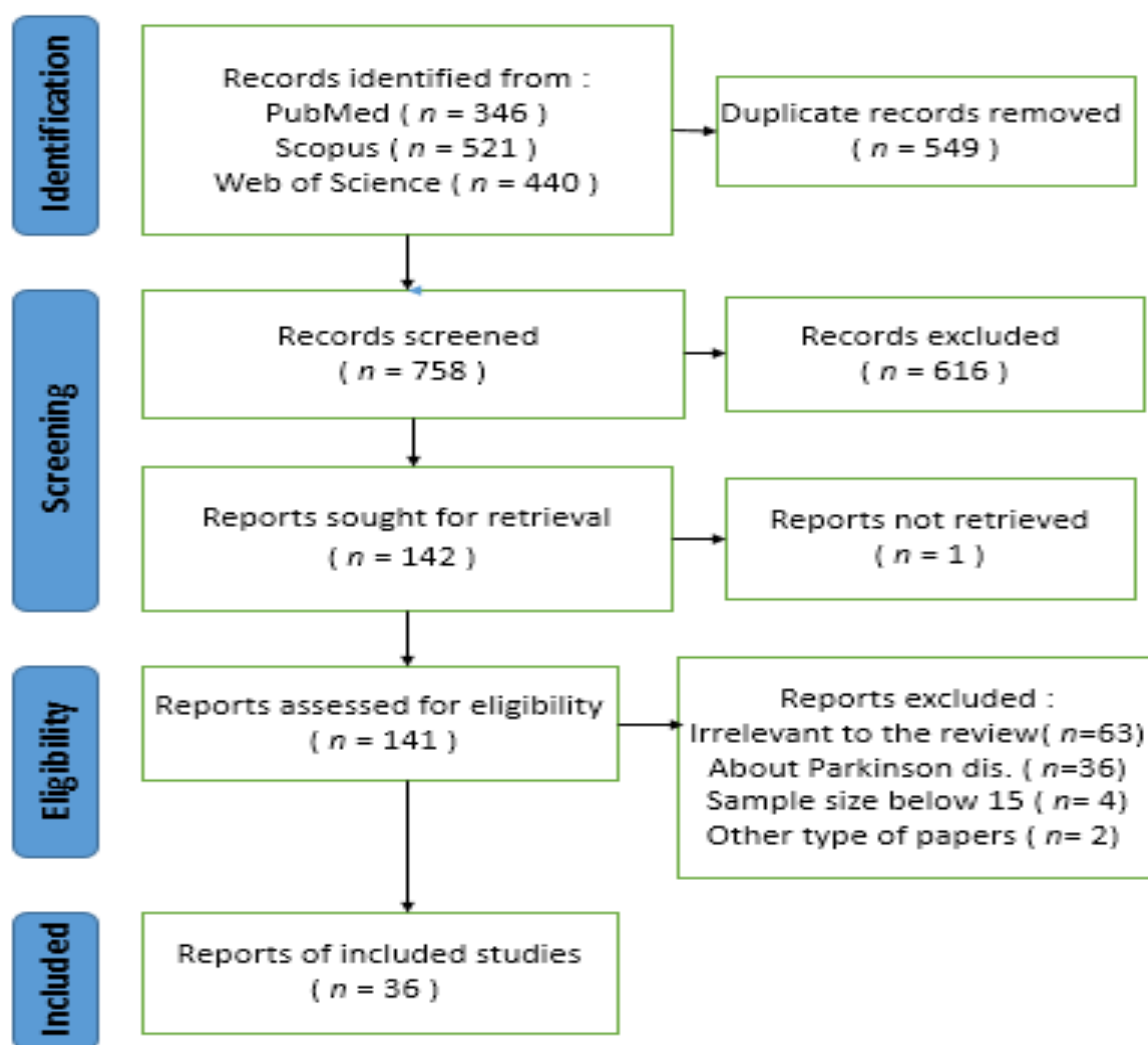
4 reports had a sample size smaller than 15 ( $n < 15$ ).

Two reports were categorized as other articles, such as literature reviews and case reports.

Inclusion Phase: Following a thorough screening and assessment process, 36 reports of eligible research were selected for further analysis and synthesis in the systematic review as shown in PRISMA chart in figure 1.

### Quality of the included studies

Upon careful examination of the studies evaluated, it can be determined that all included studies were in the third or fourth level of evidence, as per the OCEBM standards. These levels pertain to case-control studies, which are observational studies that include comparing a group of persons with a certain disease (cases) to a group without the disease (controls). (Table 1)



**Figure 1:** PRISMA chart of the included studies

#### Assessing Risk of bias

The assessment of the quality of studies using the NOS is shown in Tables 2 and 3. Four studies received a full score. Twenty (56%) of the studies fell into the good category, whereas sixteen (44%) of the studies were fair. The non-respondent rate in cross-sectional and case-control studies received the lowest scores.

#### Characteristics of the studies

The extensive data obtained from our team's research was carefully filtered according to the inclusion criteria. This table presents the salivary biomarkers for Alzheimer's disease (AD) and provides readers with a comprehensive summary of the current research on salivary biomarkers for AD. The table not only provides a summary of the key features of each study, such as the year of publication, geographical location, and demographics, but also includes important methodological details. Peña-Bautista et al., 2020 studied the minimum number, 14 AD patients <sup>(20)</sup>. While, Carro et al., 2017 studied the maximum number, 80 AD patients <sup>(21)</sup>. The studies classified according to the continents as (10) from Asia, (15) from Europe, (7) from North America, (2) from South America, (2) from Australia. Most of studies found a significant increase of salivary A $\beta$ 42 in AD patients <sup>(7,22–25)</sup> so, we can regard A $\beta$ 42 as a best marker in early AD

diagnosis by salivary sample. Most of studies used enzyme-linked immunosorbent assay (ELISA) as a method of markers determination<sup>(21–34)</sup> so, it is considered as a better method. These details are essential for evaluating and comparing the results of different studies as shown below in Table 4.

**Table 1:** Results of Oxford Centre for Evidence-Based Medicine

Study	Year	Level
Ashton et al <sup>(11)</sup>	2018	3b
Peña-Bautista et al <sup>(20)</sup>	2019	4
Carro et al <sup>(21)</sup>	2017	3b
Bermejo-Pareja et al <sup>(22)</sup>	2010	4
Santos et al <sup>(23)</sup>	2020	4
Katsipis et al <sup>(24)</sup>	2021	3b
Zalewska et al <sup>(25)</sup>	2021	3b
Lau et al <sup>(26)</sup>	2017	4
lee et al <sup>(27)</sup>	2017	3b
McGeer et al <sup>(28)</sup>	2018	4
Pukhalskaia et al <sup>(29)</sup>	2020	4
González-Sánchez et al <sup>(30)</sup>	2020	3b
Gleerup et al <sup>(31)</sup>	2021	3b
McNicholas et al <sup>(32)</sup>	2022	4
Sabaei et al <sup>(33)</sup>	2023	3b
Huan et al <sup>(34)</sup>	2018	3b
Peña-Bautista et al <sup>(35)</sup>	2020	4
Lau et al <sup>(36)</sup>	2015	3b
Shi et al <sup>(37)</sup>	2023	3b
Bakhtiari et al <sup>(38)</sup>	2017	3b
Yilmaz et al <sup>(39)</sup>	2017	3b
Sabbagh et al <sup>(40)</sup>	2018	3b
Tvarijonaviciute et al <sup>(41)</sup>	2020	3b
Boschi et al <sup>(42)</sup>	2022	3b
Eldem et al <sup>(43)</sup>	2022	3b
Marksteiner et al <sup>(44)</sup>	2022	3b
Boston et al <sup>(45)</sup>	2008	4
Souza-Talarico et al <sup>(46)</sup>	2008	4
Kim et al <sup>(47)</sup>	2014	4
Liang et al <sup>(48)</sup>	2015	4
Ahmadi-Motamayel et al <sup>(49)</sup>	2019	4
Pekeles et al <sup>(50)</sup>	2019	4
François et al <sup>(51)</sup>	2021	4
Cui et al <sup>(52)</sup>	2022	4
Ryu et al <sup>(53)</sup>	2023	4
Contini et al <sup>(54)</sup>	2023	4

**Table 2:** Assessment of case control studies involved in the systematic review using Newcastle – Ottawa quality scale.

	Year	Selection				Compa- rability	Outcome			Total
		Is the case definition adequate?	Representativeness of the cases	Selection of Controls	Definition of Controls	Comparability of cases and controls based on the design	Ascertainment of exposure	Same method of ascertainment for cases and controls	Non-Response rate	
Peña-Bautista et al <sup>(20)</sup>	2019	*	*	*	*	*	*	*		7
Bermejo-Pareja et al <sup>(22)</sup>	2010	*	*	*	*	**	*	*	*	9
Santos et al <sup>(23)</sup>	2020	*	*	*	*	*	*	*		7
Lau et al <sup>(26)</sup>	2017	*	*	*	*	*	*	*	*	8
McGeer et al <sup>(28)</sup>	2018	*	*	*	*	*	*	*		7
Pukhalskaia et al <sup>(29)</sup>	2020	*	*	*	*	*	*	*		7
McNicholas et al <sup>(32)</sup>	2022	*	*	*	*	*	*	*		7
Peña-Bautista et al <sup>(35)</sup>	2020	*	*	*	*	*	*	*		7
Boston et al <sup>(45)</sup>	2008	*	*	*	*	*	*	*	*	8
Souza-Talarico et al <sup>(46)</sup>	2008	*	*	*	*	*	*	*		7
Kim et al <sup>(47)</sup>	2014	*	*	*	*	*	*	*	*	8
Liang et al <sup>(48)</sup>	2015	*	*	*	*	*	*	*		7
Ahmadi-Motamayel et al <sup>(49)</sup>	2019	*	*	*	*	*	*	*	*	8
Pekes et al <sup>(50)</sup>	2019	*	*	*	*	*	*	*	*	9
François et al <sup>(51)</sup>	2021	*	*	*	*	**	*	*	*	9
Cui et al <sup>(52)</sup>	2022	*	*	*	*	**	*	*	*	9
Ryu et al <sup>(53)</sup>	2023	*	*	*	*	*	*	*	*	8
Contini et al <sup>(54)</sup>	2023	*	*	*	*	*	*	*	*	8

Newcastle-Ottawa scale adapted for (case control) studies; good studies: 7- 9; fair: 4-6 points; poor: 0-3 points, each category with selection and outcome can be awarded up to one star, comparability can be awarded up to two stars.

**Table 3:** Assessment of case studies involved in the systematic review using Newcastle – Ottawa quality scale.

Study	Year	Selection			Comparability	Outcome		Total
		Representativeness of the cases	Sample size	Non-Response rate		Assessment of the outcome	Statistical test:	
Ashton et al <sup>(11)</sup>	2018	*			*	*	*	5
Carro et al <sup>(21)</sup>	2017	*	*		*	*	*	6
Katsipis et al <sup>(24)</sup>	2021	*	*		*	*	*	6
Zalewska et al <sup>(25)</sup>	2021	*	*		*	*	*	6
lee et al <sup>(27)</sup>	2015	*	*	*	*	*	*	7
González-Sánchez et al <sup>(30)</sup>	2020	*	*		*	*	*	6
Gleerup et al <sup>(31)</sup>	2021	*	*		*	*	*	6
Sabaei et al <sup>(33)</sup>	2023	*	*		*	*	*	6
Huan et al <sup>(34)</sup>	2018	*	*		*	*	*	6
Lau et al <sup>(36)</sup>	2015	*	*		*	*	*	6
Bakhtiari et al <sup>(38)</sup>	2017	*	*		*	*	*	6
Yilmaz et al <sup>(39)</sup>	2017	*	*		*	*	*	6
Sabbagh et al <sup>(40)</sup>	2018	*	*		*	*	*	6
Tvari-jonaviciute et al <sup>(41)</sup>	2020	*	*		*	*	*	6
Boschi et al <sup>(42)</sup>	2022	*			*	*	*	5
Eldem et al <sup>(43)</sup>	2022	*	*	*	*	*	*	7
Marksteiner et al <sup>(44)</sup>	2022	*	*		*	*	*	6
Shi et al <sup>(55)</sup>	2011	*	*		*	*	*	6

Newcastle-Ottawa scale adapted for cross-sectional studies; good studies: 7- 9; fair: 4-6 points; poor: 0-3 points



**Table 4:** Characteristics of included studies

Author, Year	Study Group (F/M), Age (year )	Control Group, (F/M), Age (year)	Method of Marker Determination	Salivary Biomarkers
<b>Ashton et al., 2018</b> <sup>(11)</sup>	AD: 53 (30/23), (81.4 ± 6.6); MCI: 68 (35/33), (79.8 ± 6.4)	160 (94/66), (78.0 ± 6.7)	SIMOA	t-tau (NS)
<b>Peña-Bautista et al 2019</b> <sup>(20)</sup>	mild AD: 50 (29/21), 70(68-74); MCI-AD: 47 (30/17), 71(69-74)	41 (16/25), 66 (62-69)	UPLC-MS/MS	cortisol (NS)
<b>Carro et al., 2017</b> <sup>(21)</sup>	AD: 80 (49/31), (76.2 ± 5.33) ; MCI: 44 (25/19), (75.16 ± 5.13)	91 (59/32), (73.7 ± 6.88)	ELISA	lactoferrin
<b>Bermejo-Pareja et 2010</b> <sup>(22)</sup>	AD: 70 (49/21), 77.20(60-91)	56 (39/17), 74.35(64-85)	ELISA	Aβ42, Aβ40 (NS) (beta-amyloid)
<b>Santos et al., 2020</b> <sup>(23)</sup>	AD: 60 (NR), NR	60 (NR), NR	ELISA	Aβ42, t-tau
<b>Katsipis et al., 2021</b> <sup>(24)</sup>	AD: 20 (9/11), (75 ± 5.5); MCI: 20 (12/8), (75 ± 7.2)	20 (11/9), (79 ± 4.7)	ELISA, Dot Blot, Western Blot	GFAP, p-tau, IL-1β, IL-6, TNF-α, caspase-8, COX-2, Aβ42
<b>Zalewska et al., 2021</b> <sup>(25)</sup>	AD: 25 (15/10), (81.19 ± 6.77)	25 (15/10), (82.1 ± 6.67)	colorimetric, spectrofluorimetric, spectrophotometric methods, thioflavin T fluorescence, ELISA	Aβ, lactoferrin, IL-1β, SOD, CAT, GPx, UA (NS), GSH, TAC (NS), TOS, AGE, AOPP, MDA, NO, peroxytrite, nitrotyrosine
<b>Lau et al., 2017</b> <sup>(26)</sup>	AD: 28, (76.4±1.8)	17, (63.4±1.6)	EG-IDFET biosensor	Trehalose
<b>Lee et al., 2017</b> <sup>(27)</sup>	AD: 28, (77.3±1.3)	17, (67.8±1.4)	ELISA	Aβ42
<b>McGeer et al., 2018</b> <sup>(28)</sup>	AD: 27, (83.0±1.5)	28, (68.0±1.3)	ELISA	Aβ42
<b>Pukhalskaia et al., 2020</b> <sup>(29)</sup>	AD: 64 (NR), elderly (63.0 ± 2.4), senile (82.0 ± 2.3)	58 (NR), NR	ELISA	SIRT1, SIRT3, SIRT5 (NS), SIRT6
<b>González-Sánchez al., 2020</b> <sup>(30)</sup>	AD-PET+: 25 (12/13), (67.2 ± 9.2), MCI-PET+: 21 (8/13), (68.8 ± 7.5)	Control-PET-: 48 (33/15), 66.9 ± 5.9; control-PET+: 4 (2/2), (75.9 ± 3.6)	ELISA	lactoferrin
<b>Gleerup et al., 2021</b> <sup>(31)</sup>	AD: 71 (41/30), (72.1 ± 7.3); MCI: 56 (27/29), (70.4 ± 8.2)	20 (8/12), (65.7 ± 10.1)	ELISA	lactoferrin
<b>McNicholas et al., 2022</b> <sup>(32)</sup>	AD: 16 (6/10), (79 ± 6); MCI: 15 (8/7), (76 ± 6)	29 (14/15), (74 ± 8)	ELISA	cystatin-C, IL-1 receptor antagonist, stratifin, haptoglobin, matrix metalloproteinase 9
<b>Sabaei et al., 2023</b> <sup>(33)</sup>	AD: 24 (10/14), (73.5 ± 9.8)	22 (13/9), (64.1 ± 9.2)	ELISA	Aβ 1--42, p-tau, α-synuclein
<b>Huan et al., 2018</b> <sup>(34)</sup>	AD: 22 (16/6), (77.09 ± 11.20); MCI: 25 (15/10), (70.40 ± 3.38)	35 (22/13), (69.94 ± 3.80); 10 (5/5), (71.40 ± 2.84)	LC-MS	metabolomics
<b>Peña-Bautista et al 2020</b> <sup>(35)</sup>	mild AD: 14 (NR), NR; MCI-AD: 17 (NR), NR	12 (NR), NR	UPLC-MS/MS	aspartic acid (NS), glutamic acid (NS), glutamine, GABA (NS), creatine, taurine, N-acetyl aspartate (NS), myoinositol, acetylcholine
<b>Lau et al., 2015</b> <sup>(36)</sup>	AD: 20 (12/8), (72.5 ± 7.68)	20 (15/5), (66.1 ± 7.79)	ELISA, EG-ISFET	Aβ42 (not detected), p-tau (NS), t-tau (NS), trehalose
<b>Bakhtiari et al., 2017</b> <sup>(38)</sup>	AD: 15 (6/9), 78.4 (64-90)	15 (8/7), 71 (61-85)	Ellman colorimetric method	AChE (NS)

<b>Yilmaz et al., 2017</b> <sup>(39)</sup>	AD: 22, (70.5±1.8)	35, (69.7±1.2)	Proton NMR spectroscopy	Propionate
<b>Sabbagh et al., 2018</b> <sup>(40)</sup>	AD: 15, (77.8±1.8)	7, (70.4±3.5)	ELISA	Aβ42
<b>Tvarijonavičiute et al., 2020</b> <sup>(41)</sup>	AD: 69 (NR), NR	83 (NR), NR	MILLIPLEX MAP, automated spectrophotometric method	FRAP (NS), ADA (NS), ChE (NS), Hp (NS), Aβ42, Aβ40 (NS), t-tau (NS), p-tau (NS), CRP (NS), PEDF (NS), SAP (NS), MIP-4 (NS), CC4, α1 antitrypsin (NS)
<b>Boschi et al., 2022</b> <sup>(42)</sup>	AD: 18 (10/8), (72.13 ± 5.45)	18 (11/7), (65.67 ± 12.02)	ELISA	Aβ42
<b>Eldem et al., 2022</b> <sup>(43)</sup>	AD: 17 (9/8), (72 ± 1.36); MCI: 21 (16/5), (73 ± 1.51)	19 (13/6), (64 ± 2.63)	LC-MS/MS, Western Blot, FASP	proteomics, t-tau (NS), transthyretin
<b>Marksteiner et al., 2022</b> <sup>(44)</sup>	AD: 44 (25/19), (79 ± 1); MCI: 45 (25/20), (74 ± 1)	27 (14/13), (71 ± 1)	HPLC-EC, robotic-automated enzymatic Lumipulse assay	norepinephrine, p-tau (NS), t-tau, Aβ40 and Aβ42 (not detected in cases)
<b>Boston et al., 2008</b> <sup>(45)</sup>	AD: 15, (77.6±6.6)	13, (68.8±9.1)	Ellman's colorimetric method	AChE activity
<b>Souza-Talarico et al., 2008</b> <sup>(46)</sup>	AD:40 (27/13), (80.1 ± 6.0)	40 (35/5), (72.2 ± 6.3)	Radioimmunoassay	Cortisol
<b>Kim et al., 2014</b> <sup>(47)</sup>	AD: 28, (66-88 )	17, (65-87 )	Immunoassay with nanobeads	Aβ42, Aβ40
<b>Liang et al., 2015</b> <sup>(48)</sup>	AD: 256 (NR)	218 (NR)	FUPLC-MS	metabolomics
<b>Ahmadi-Motamay et al., 2019</b> <sup>(49)</sup>	AD: 30 (NR), (70.57±6.317)	30 (NR), (66.50±8.046)	Ellman colorimetric method	AChE, PChE (Pseudo-cholinesterase)
<b>Pekales et al., 2019</b> <sup>(50)</sup>	AD: 46 (22/24), (80 ±9); MCI: 55 (32/23), (78 ±14)	47 (32/15), (73 ±6)	Western Blot, tau antibodies	p-tau (T181 (NS), S396, S404, S400, T403, T404)
<b>François et al., 2021</b> <sup>(51)</sup>	AD: 20 (8/12), 78.0; MCI: 20 (11/9), 77.8	40 (19/21), 75.3	GC-MS, LC-MS	proteomics, metabolomics
<b>Cui et al., 2022</b> <sup>(52)</sup>	AD: 30 (NR), NR	30 (NR), NR	-	Aβ42, Aβ40 (NS)
<b>Ryu et al., 2023</b> <sup>(53)</sup>	AD: 27 (12/15), (72.59 ± 6.90)	13 (11/2), (75.46 ± 6.58)	qPCR	miRNA-485-3p
<b>Contini et al., 2023</b> <sup>(54)</sup>	AD: 35 (23/12), (80 ± 6)	36 (18/18), (78 ± 6)	RP-HPLC-LR-ESI-MS	proteomics
<b>Shi et al., 2011</b> <sup>(55)</sup>	AD: 21 (11/10)	38 (19/19)	Luminex assays, IP/MS	t-tau, p-tau, Aβ42 (not detected)

Abbreviations: Aβ, beta-amyloid; AD, Alzheimer's Disease; ChE, cholinesterase; COX-2, cyclooxygenase 2; CRP, C-reactive protein; ELISA, enzyme-linked immunosorbent assay; GFAP, glial fibrillar acidic protein; LF, lactoferrin; MCI, mild cognitive impairment; NO, nitric oxide; NR, not reported; ns, not significant; OSI, oxidative stress index; p-tau, phosphorylated tau; qPCR, quantitative real-time polymerase chain reaction; SOD, superoxide dismutase; t-tau, total tau

## Discussion

Based on the comprehensive analysis of this subject, it can be seen that some biomarkers have the ability to be identified in the saliva of individuals with Alzheimer's disease. This systematic review revealed a significant increase in beta-amyloid42 and phosphorylated tau (p Tau) levels in people with Alzheimer's disease compared to healthy individuals <sup>(1)</sup>. However, there was a slight decrease in total tau and lactoferrin levels, but this difference did not achieve statistical significance <sup>(56)</sup>. Moreover, the histopathological features of AD such as beta-amyloid plaques and hyperphosphorylated tau tangles are found to be in individual brain samples. Saliva contains the primary indicators for Alzheimer's disease (AD), suggesting that the development of non-invasive diagnostic procedures has great promise.

## Beta-amyloid

The peptide which is mainly found in neurons and that can be present in pathological plaques which can be characterized as hallmark of AD is called beta-amyloid (A $\beta$ )<sup>(57)</sup>.

Many studies were in line with a significant increase of salivary A $\beta$ 42 in AD patients,<sup>(22–25,33,42)</sup> while others did not find such alteration<sup>(36,44)</sup>. Bermejo-Pareja et al. (2010) discovered that individuals with mild Alzheimer's disease (AD) had significantly elevated levels of salivary A $\beta$ 42 compared to the control group. They also observed a similar but non-significant trend in patients with moderate and severe AD<sup>(22)</sup>. The levels of A $\beta$ 42 showed a correlation with sex, whereas A $\beta$ 40 did not exhibit such correlation. In their study, Santos et al. (2020) found that there was a little increase in salivary A $\beta$ 42 levels in patients with Alzheimer's disease, however this increase was not statistically significant<sup>(23)</sup>. Cui et al. (2022) discovered a notable rise in A $\beta$ 42 levels, but not A $\beta$ 40 levels, in the saliva of individuals with Alzheimer's disease (AD). However, neither of these biomarkers alone demonstrated predictive capability<sup>(52)</sup>. Katsipis et al. (2021) and Boschi et al. (2022) found that individuals with Alzheimer's disease (AD) had noticeably higher levels of salivary A $\beta$ 42 compared to individuals without AD. This increase was observed regardless of gender or MMSE score<sup>(24,42)</sup>. Salivary A $\beta$ 42 had good diagnostic performance (AUC 0.806) With a cut-off value of 92.5 pg/mL, sensitivity and specificity are 0.84 and 0.68, respectively in Boschi et al. (2022) study.<sup>(42)</sup> In another study, Sabaei et al. (2023) found that individuals with Alzheimer's disease (AD) had significantly elevated levels of salivary A $\beta$ 1–42, even after accounting for age differences, Salivary A $\beta$  1–42 level at a 60.3 pg/ml cutoff point revealed an excellent performance for diagnosing AD (AUC: 0.81)<sup>(33)</sup>. Contrarily, Tvarijonaviciute et al. (2020) discovered a reduction in salivary A $\beta$ 42 levels in people with Alzheimer's disease (AD) when compared to individuals without the condition. This decrease was observed regardless of the stage of the disease, and no correlation was established between A $\beta$ 42 levels and the progression of the disease<sup>(41)</sup>. Zalewska et al. (2021) used thioflavin T fluorescence to evaluate the  $\beta$ -amyloid formation (The intensity of the amyloid associated with thioflavin T fluorescence was measured at 385/485 nm wavelength) and found significantly higher salivary A $\beta$  in AD (AUC 0.949)<sup>(25)</sup>. Nevertheless, Lau et al. (2015), Marksteiner et al. (2022), and Shi et al. (2011) were unable to identify the presence of salivary A $\beta$ 40 or A $\beta$ 42 in individuals with Alzheimer's disease using ELISA, automated enzymatic tests, or Luminex technology<sup>(36,44,55,58)</sup>.

## Lactoferrin

Lactoferrin is an iron-binding protein with potential neuroprotective effects in AD<sup>(59)</sup>. Lactoferrin, a protein with neuroprotective effects and serving as a biomarker, has recently shown potential as a compelling candidate warranting additional investigation<sup>(59)</sup>. Most studies found significantly decreased salivary lactoferrin in AD and MCI patients compared to controls,<sup>(21,25,30)</sup> although one study found no difference<sup>(31)</sup>. Carro et al. (2017) reported significantly lower salivary lactoferrin in AD and MCI compared to controls, correlating with disease severity and MMSE scores. Lactoferrin had perfect diagnostic accuracy (AUC 1.0)<sup>(21)</sup>. González-Sánchez et al. (2020) found decreased lactoferrin in MCI and AD patients with positive amyloid PET scans versus controls (AUC 0.952) using the Youden's index, found that the optimal cut-off point to differentiate salivary Lactoferrin from MCI/AD-PET+patients and control-PET group was 5.63mg/ml<sup>(30)</sup>. Zalewska et al. (2021) also observed significantly reduced salivary lactoferrin in AD (AUC 0.690)<sup>(25)</sup>. However, Gleerup et al. (2021) found no significant difference and a non-significant trend for increased lactoferrin in AD saliva, possibly due to including milder, more heterogeneous cases<sup>(31)</sup>.

## Acetylcholinesterase and Related Enzyme

Acetylcholinesterase (AChE) breaks down the neurotransmitter acetylcholine and is a key AD treatment target <sup>(60,61)</sup>. Two studies reported conflicting results on salivary AChE activity in AD <sup>(38,49)</sup>. Pseudocholinesterase and general cholinesterase activity findings were also mixed <sup>(41,49)</sup>. Ahmadi-Motamayel et al. (2019) found significantly increased AChE and pseudocholinesterase activity in AD saliva, more so in males <sup>(49)</sup>. However, Bakhtiari et al. (2017) reported non-significantly decreased AChE activity in AD patients on memantine therapy <sup>(38)</sup>. Tvarijonaviciute et al. (2020) observed a non-significant trend for increased cholinesterase in AD saliva <sup>(41)</sup>.

## Tau Protein

Tau proteins stabilize neuronal microtubules but can become hyperphosphorylated and aggregate in AD, impairing synaptic function <sup>(62,63)</sup>. Studies found mixed results for salivary total tau (t-tau) and phosphorylated tau (p-tau) in AD. Shi et al. (2011) reported non-significantly decreased salivary t-tau and increased p-tau in AD, with a significantly higher p-tau/t-tau ratio <sup>(55)</sup>.

Lau et al. (2015) detected p-tau and t-tau but found no differences between AD and controls <sup>(36)</sup>. Ashton et al. (2018) found a non-significant trend for increased salivary t-tau in AD, correlating with poorer cognition <sup>(11)</sup>. Cui et al. (2022) observed no relationship between p-tau and t-tau, but the p-tau/t-tau ratio was significantly higher in AD. Combining p-tau, t-tau, A $\beta$ 40 and A $\beta$ 42 had excellent diagnostic value (AUC 0.921) <sup>(52)</sup>. Santos et al. (2020) and Marksteiner et al. (2022) both reported significantly decreased salivary t-tau in AD patients <sup>(23,44)</sup>. However, Eldem et al. (2022) found no difference in t-tau levels between AD, MCI and controls using Western blot.<sup>(43)</sup> Marksteiner et al. (2022), and Sabaei et al. (2023) found significantly increased salivary p-tau in AD compared to controls, with good diagnostic performance <sup>(33,44)</sup>. Marksteiner et al. (2022) found laboratory diagnosis with a cut-off of  $\geq 18$  pg/mg protein pTau181 (for MCI) and  $\leq 300$  pg/mg protein tau (for AD) with an accuracy of 71.4% <sup>(44)</sup>. Sabaei et al. (2023) found the cut-off point of p-tau in AD patients equal to 5.1 pg/ml with (AUC: 0.78) <sup>(33)</sup>. Pেকেles et al. (2019) reported significantly higher p-tau/t-tau ratios at specific phosphorylation sites in AD saliva.<sup>(50)</sup> However, Tvarijonaviciute et al. (2020) found no significant differences in salivary p-tau or t-tau between AD and controls <sup>(41)</sup>.

## Inflammation and Oxidative Stress Markers

AD involves chronic inflammation and oxidative damage <sup>(4,64)</sup>. Studies found altered levels of various salivary inflammatory and oxidative stress biomarkers in AD, such as haptoglobin, interleukins, TNF-alpha, antioxidant enzymes, and oxidative damage products <sup>(24,32,41)</sup>.

Tvarijonaviciute et al. (2020) observed decreased haptoglobin, adenosine deaminase, and ferric-reducing ability of plasma, and increased macrophage inflammatory protein-4,  $\alpha$ 1-antitrypsin, complement C4, and pigment epithelium-derived protein in AD saliva, with only complement C4 being significant <sup>(41)</sup>. Katsipis et al. (2021) found significantly decreased glial fibrillary acidic protein (GFAP), COX-2, and caspase-8, and increased IL-1 $\beta$ , IL-6, and TNF- $\alpha$  in AD saliva compared to MCI and controls. These markers had excellent diagnostic accuracy (AUC up to 1.0) and correlated with each other and MMSE scores. The proposed ELISA cut-off values, determined using the J indexes for various thresholds, are as follows: for cognitively healthy individuals, [GFAP] levels exceed 10.0 ng/mg; for MCI patients, [GFAP] levels range between 4.0 and 10.0 ng/mg (J index = 0.95, 95% sensitivity, and 100% specificity); and for AD patients, [GFAP] levels fall below 4.0 ng/mg of total protein (J index = 0.75, 75% sensitivity, and 100% specificity <sup>(24)</sup>. Zalewska et

al. (2021) reported significantly reduced antioxidant enzymes (catalase, glutathione peroxidase, superoxide dismutase) and glutathione, and increased oxidative damage markers (nitric oxide, protein oxidation products, lipid peroxidation, interleukin-1 $\beta$ ) in stimulated AD saliva. Many had good diagnostic performance like nitric oxide (AUC 0.672) with cut-off value > 0.1961 ng/mg <sup>(25)</sup>. McNicholas et al. (2022) found altered levels of cystatin-C, IL-1 receptor antagonist, stratifin, haptoglobin and matrix metalloproteinase 9 in AD and MCI saliva. Panels combining age, sex, APOE status and some of these markers adjusted for total protein had excellent accuracy to distinguish AD and MCI from controls (AUC 0.97) <sup>(32)</sup>.

### Cortisol

Cortisol is a stress hormone that may contribute to AD pathology <sup>(27,65)</sup>. One study found increased salivary cortisol in mild AD, <sup>(46)</sup> while another found no significant difference <sup>(20,66)</sup>. De Souza-Talarico et al. (2008) reported significantly higher salivary cortisol in mild AD patients compared to controls, with a non-significant trend correlating with worse working memory <sup>(46)</sup>. However, Peña-Bautista et al. (2019) found no significant association between AD and salivary cortisol despite a non-significant increase.

### Amino Acids and Derivatives

Altered levels of amino acids and derivatives in AD saliva may reflect impaired neurotransmission. <sup>(67,68)</sup> Peña-Bautista et al. (2020) found significantly increased acetylcholine in mild AD, and decreased creatine and myoinositol in AD saliva compared to controls <sup>(35)</sup>. These correlated with cognitive scores and combining them had good diagnostic accuracy (AUC 0.806). Marksteiner et al. (2022) reported significantly decreased salivary norepinephrine in AD patients <sup>(44)</sup>.

### miRNAs and Sirtuins

MicroRNAs regulate gene expression and sirtuins are neuroprotective enzymes <sup>(69,70)</sup>. Ryu et al. (2023) found significantly increased salivary exosomal miRNA-485-3p in AD, with good diagnostic accuracy (AUC 0.895) and correlation to amyloid PET <sup>(53)</sup>. Pukhalskaia et al. (2020) reported significantly decreased salivary sirtuins 1, 3 and 6 in AD compared to controls, especially in older subjects <sup>(71)</sup>.

### Trehalose

Trehalose is a disaccharide with potential neuroprotective effects <sup>(72)</sup>. Lau et al. (2017) found increased salivary trehalose in AD using a novel biosensor method <sup>(26)</sup>.

### Metabolomics and Proteomics

Metabolomic and proteomic approaches can identify disease biomarker panels <sup>(73)</sup>. Huan et al. (2018) developed salivary metabolite panels that distinguished AD from controls and MCI with excellent accuracy (AUC >0.99) <sup>(34)</sup>. François et al. (2021) identified 79 salivary metabolites and 346 proteins significantly altered in AD versus control <sup>(51)</sup>.

### Limitations

This systematic review had certain limitations, such as the presence of variability among the included papers regarding diagnostic methodologies, inclusion/exclusion criteria, and the specific biomarkers ex-

amined. There is a requirement for standardized techniques to be established for the collection and processing of saliva in order to minimize variability. Given the relatively small sample sizes in most of the researches (26,32,34,35,39,42,46,48,53,54) it is necessary to validate the findings in bigger cohorts that are well-characterized.

## Conclusion

This systematic review indicated that AD patients have changed salivary biomarkers, namely beta-amyloid42, tau proteins, and lactoferrin. AD was associated with increased salivary beta-amyloid42 and phosphorylated tau, while decreased total tau and lactoferrin were not significant. Most of studies found a significant increase of salivary A $\beta$ 42 in AD patients. so, we can regard A $\beta$ 42 as a best marker in early AD diagnosis by salivary sample. However, salivary testing for early AD diagnosis requires high-quality research to validate these findings in bigger cohorts and standardize techniques. Biomarker panels and potential indicators such microRNAs, metabolites, and inflammatory mediators may also be beneficial. This review shows saliva's promising potential as a non-invasive AD biomarker source, despite its limits and need for more study. Salivary tests may help diagnose this terrible neurological illness earlier and more easily with current study. These tests can help track disease development and assess new disease-modifying medications. The use of salivary biomarkers in AD diagnosis may improve early detection and treatment.

## Conflict of interest

The authors have no conflicts of interest to declare.

## Author contributions

Conceptualization: NHA & AMA; Design of the study: AAM & AMA; Data Collection: AAM & NHA; Data Analysis: AAM & NHA; Critical Review: NHA &AMA; Drafting the paper: AMA & NHA

## Acknowledgement and funding

The authors would like to thank the librarian of College of Dentistry, University of Baghdad for granting access to journal's databases.

## Informed consent

NA

## References

1. Guo T, Zhang D, Zeng Y, Huang TY, Xu H, Zhao Y. Molecular and cellular mechanisms underlying the pathogenesis of Alzheimer's disease. *Mol Neurodegener.* 2020;15(1):40. <https://doi.org/10.1186/s13024-020-00391-7>
2. Zhang XX, Tian Y, Wang ZT, Ma YH, Tan L, Yu JT. The epidemiology of Alzheimer's disease modifiable risk factors and prevention. *J Prev Alzheimers Dis.* 2021;8(3):313–21. <https://doi.org/10.14283/jpad.2021.15>
3. Porsteinsson AP, Isaacson RS, Knox S, Sabbagh MN, Rubino I. Diagnosis of early Alzheimer's disease: clinical practice in 2021. *J Prev Alzheimers Dis.* 2021;8(3):371–86. <https://doi.org/10.14283/jpad.2021.23>

4. Chen Z, Zhong C. Oxidative stress in Alzheimer's disease. *Neurosci Bull.* 2014;30(2):271–81. . <https://doi.org/10.1007/s12264-013-1423-y>
5. Song T, Song X, Zhu C, Patrick R, Skurla M, Santangelo I, et al. Mitochondrial dysfunction, oxidative stress, neuroinflammation, and metabolic alterations in the progression of Alzheimer's disease: A meta-analysis of in vivo magnetic resonance spectroscopy studies. *Ageing Res Rev.* 2021; 72:101503. <https://doi.org/10.1016/j.arr.2021.101503>
6. Lane CA, Hardy J, Schott JM. Alzheimer's disease. *Eur J Neurol.* 2018;25(1):59–70. <https://doi.org/10.1111/ene.13439>
7. Mendiola-Precoma J, Berumen LC, Padilla K, Garcia-Alcocer G. Therapies for prevention and treatment of Alzheimer's disease. *Biomed Res Int.* 2016;2016(1):2589276. <https://doi.org/10.1155/2016/2589276>
8. Jack Jr CR, Bennett DA, Blennow K, Carrillo MC, Dunn B, Haeberlein SB, et al. NIA-AA research framework: toward a biological definition of Alzheimer's disease. *Alzheimer's & dementia.* 2018;14(4):535–62. <https://doi.org/10.1016/j.jalz.2018.02.018>
9. Mahaman YAR, Embaye KS, Huang F, Li L, Zhu F, Wang JZ, et al. Biomarkers used in Alzheimer's disease diagnosis, treatment, and prevention. *Ageing Res Rev.* 2022; 74:101544. <https://doi.org/10.1016/j.arr.2021.101544>
10. Ibrahim AA, Ai-Lami F, Al-Rudainy R, Khader YS. Mental disorders among elderly people in Baghdad, Iraq, 2017. *INQUIRY: The Journal of Health Care Organization, Provision, and Financing.* 2019; 56. <https://doi.org/10.1177/0046958019845960>
11. Ashton NJ, Ide M, Zetterberg H, Blennow K. Salivary biomarkers for Alzheimer's disease and related disorders. *Neuro. and Ther.* 2019 Dec; 8:83-94. <https://doi.org/10.1117/1.JBO.19.5.051205>
12. Hussein SI, Abd ST, AL-khayat FA, k Mahmood H. Complete blood count and saliva parameters as an indicator for infected patients with coronavirus COVID-19. *J Bagh Coll Den.* 2023;35(1):76–85. <https://doi.org/10.26477/jbcd.v35i1.3317>
13. Al-Joboury AAM, Rasheed RH. An Assessment of Oral Health in Hypertensive Patients Treated with HMG-CoA Reductase Inhibitors (Statins). *J Bagh Coll Dent.* 2015;27(4):85–9.
14. Al-Kubaisi MW, Al-Ghurabi BH, Alkubaisy W, ABDULLAH NN. Anti-inflammatory effects of manuka honey on salivary cytokines (clinical study). *J Bagh Coll Dent.* 2023;35(1):10–9. <https://doi.org/10.26477/jbcd.v35i1.3310>
15. Gul SS, Abdulkareem AA, Sha AM, Rawlinson A. Diagnostic accuracy of oral fluids biomarker profile to determine the current and future status of periodontal and peri-implant diseases. *Diagnostics.* 2020;10(10):838. <https://doi.org/10.3390/diagnostics10100838>
16. Al-Taweel FBH, Saliem SS, Abd OH, Whawell SA. Assessment of serum interleukin-1 $\beta$  and interleukin-6 levels in patients with chronic periodontitis and coronary heart disease. *European J Gen Dent.* 2021;10(02):78–83. <https://doi.org/10.1055/s-0041-1732954>
17. Page MJ, McKenzie JE, Bossuyt PM, Boutron I, Hoffmann TC, Mulrow CD, et al. The PRISMA 2020 statement: an updated guideline for reporting systematic reviews. *bmj.* 2021;372. . <https://doi.org/10.1136/bmj.n71>
18. OCEBM Levels of Evidence Working Group. The Oxford 2011 Levels of Evidence. Oxford Centre for Evidence-Based Medicine. <http://www.cebm.net/index.aspx?o=5653>. Accessed 22 Sept 2012
19. Wells GASB, O'Connell D, Peterson J, Welch V, Losos M, Tugwell P. The Newcastle -Ottawa scale (NOS) for assessing the quality of nonrandomized studies in meta-analyses. 2011

20. Peña-Bautista C, Baquero M, Ferrer I, Hervás D, Vento M, García-Blanco A, et al. Neuropsychological assessment and cortisol levels in biofluids from early Alzheimer's disease patients. *Exp Gerontol.* 2019; 123:10–6. <https://doi.org/10.1016/j.exger.2019.05.007>
21. Carro E, Bartolomé F, Bermejo-Pareja F, Villarejo-Galende A, Molina JA, Ortiz P, et al. Early diagnosis of mild cognitive impairment and Alzheimer's disease based on salivary lactoferrin. *Alzheimer's & Dementia: Diagnosis, Assessment & Disease Monitoring.* 2017; 8:131–8. <https://doi.org/10.1016/j.dadm.2017.04.002>
22. Bermejo-Pareja F, Antequera D, Vargas T, Molina JA, Carro E. Saliva levels of Abeta1-42 as potential biomarker of Alzheimer's disease: a pilot study. *BMC Neurol.* 2010;10(1):108. <https://doi.org/10.1186/1471-2377-10-108>
23. Santos GAA, Olave E, Pardi PC. Salivary Biomarkers in Alzheimer's Disease. *International Journal of Morphology.* 2020;38(1):230-4. <https://doi.org/10.31053/1852.4834.v38.n1.2853>
24. Katsipis G, Tzekaki EE, Tsolaki M, Pantazaki AA. Salivary GFAP as a potential biomarker for diagnosis of mild cognitive impairment and Alzheimer's disease and its correlation with neuroinflammation and apoptosis. *J Neuroimmunol.* 2021; 361:577744. <https://doi.org/10.1016/j.jneuroim.2021.577744>
25. Zalewska A, Klimiuk A, Zięba S, Wnorowska O, Rusak M, Waszkiewicz N, et al. Salivary gland dysfunction and salivary redox imbalance in patients with Alzheimer's disease. *Sci Rep.* 2021;11(1):23904. <https://doi.org/10.1038/s41598-021-03456-9>
26. Lau HC, Yu JB, Lee HW, Huh JS, Lim JO. Investigation of exhaled breath samples from patients with Alzheimer's disease using gas chromatography-mass spectrometry and an exhaled breath sensor system. *Sensors.* 2017;17(8):1783. . <https://doi.org/10.3390/s17081783>
27. Lee DY, Kim E, Choi MH. Technical and clinical aspects of cortisol as a biochemical marker of chronic stress. *BMB Rep.* 2015;48(4):209-16. <https://doi.org/10.5483%2FBMBRep.2015.48.4.275>
28. McGeer PL, Guo JP, Lee M, Kennedy K, McGeer EG. Alzheimer's disease can be spared by nonsteroidal anti-inflammatory drugs. *J Alzheimer's Dis.* 2018;62(3):1219–22. <https://doi.org/10.3233/JAD-180614>
29. Pukhalskaia AE, Dyatlova AS, Linkova NS, Kozlov KL, Kvetnaia T V, Koroleva M V, et al. Sirtuins as possible predictors of aging and Alzheimer's disease development: verification in the hippocampus and saliva. *Bull Exp Biol Med.* 2020;169(6):821–4. <https://doi.org/10.1007/s10517-020-04986-4>
30. González-Sánchez M, Bartolome F, Antequera D, Puertas-Martín V, González P, Gómez-Grande A, et al. Decreased salivary lactoferrin levels are specific to Alzheimer's disease. *EBioMedicine.* 2020;57. <https://doi.org/10.1016/j.ebiom.2020.102834>
31. Gleerup HS, Jensen CS, Høgh P, Hasselbalch SG, Simonsen AH. Lactoferrin in cerebrospinal fluid and saliva is not a diagnostic biomarker for Alzheimer's disease in a mixed memory clinic population. *EBioMedicine.* 2021;67. <https://doi.org/10.1016/j.ebiom.2021.103361>
32. McNicholas K, François M, Liu JW, Doecke JD, Hecker J, Faunt J, et al. Salivary inflammatory biomarkers are predictive of mild cognitive impairment and Alzheimer's disease in a feasibility study. *Front Aging Neurosci.* 2022; 14:1019296. <https://doi.org/10.3389/fnagi.2022.1019296>



33. Sabaei M, Rahimian S, Ketabforoush AHME, Rasoolijazi H, Zamani B, Hajiakhoundi F, et al. Salivary levels of disease-related biomarkers in the early stages of Parkinson's and Alzheimer's disease: a cross-sectional study. *IBRO Neurosci Rep.* 2023; 14:285–92. <https://doi.org/10.1016/j.ibneur.2023.03.004>
34. Huan T, Tran T, Zheng J, Sapkota S, MacDonald SW, Camicioli R, et al. Metabolomics analyses of saliva detect novel biomarkers of Alzheimer's disease. *J Alzheimer's Dis.* 2018;65(4):1401–16. <https://doi.org/10.3233/JAD-180351>
35. Peña-Bautista C, Torres-Cuevas I, Baquero M, Ferrer I, García L, Vento M, et al. Early neurotransmission impairment in non-invasive Alzheimer Disease detection. *Sci Rep.* 2020;10(1): 16396. <https://doi.org/10.1038/s41598-020-73362-z>
36. Lau HC, Lee IK, Ko PW, Lee HW, Huh JS, Cho WJ, et al. Non-invasive screening for Alzheimer's disease by sensing salivary sugar using *Drosophila* cells expressing gustatory receptor (Gr5a) immobilized on an extended gate ion-sensitive field-effect transistor (EG-ISFET) biosensor. *PLoS One.* 2015;10(2): e0117810. <https://doi.org/10.1371/journal.pone.0117810>
37. Shi Y, Wang H, Zhu Z, Ye Q, Lin F, Cai G. Association between exposure to phenols and parabens and cognitive function in older adults in the United States: A cross-sectional study. *Science of The Total Environment.* 2023; 858:160129. <https://doi.org/10.1016/j.scitotenv.2022.160129>
38. Bakhtiari S, Moghadam NB, Ehsani M, Mortazavi H, Sabour S, Bakhshi M. Can salivary acetylcholinesterase be a diagnostic biomarker for Alzheimer? *J Clin Diagn Res.* 2017;11(1): ZC58. <https://doi.org/10.7860%2FJCDR%2F2017%2F21715.9192>
39. Yilmaz A, Geddes T, Han B, Bahado-Singh RO, Wilson GD, Imam K, et al. Diagnostic biomarkers of Alzheimer's disease as identified in saliva using <sup>1</sup>H NMR-based metabolomics. *J Alzheimer's Dis.* 2017;58(2):355–9. . <https://doi.org/10.3233/JAD-160772>
40. Sabbagh MN, Shi J, Lee M, Arnold L, Al-Hasan Y, Heim J, et al. Salivary beta amyloid protein levels are detectable and differentiate patients with Alzheimer's disease dementia from normal controls: Preliminary findings. *BMC Neurol.* 2018;18(1):155. <https://doi.org/10.1186/s12883-018-1160-y>
41. Tvarijonaviciute A, Zamora C, Ceron JJ, Bravo-Cantero AF, Pardo-Marin L, Valverde S, et al. Salivary biomarkers in Alzheimer's disease. *Clin Oral Investig.* 2020;24(10):3437–44. <https://doi.org/10.1007/s00784-020-03214-7>
42. Boschi S, Roveta F, Grassini A, Marcinnò A, Cermelli A, Ferrandes F, et al. A $\beta$ 42 as a biomarker of Alzheimer's disease: is saliva a viable alternative to cerebrospinal fluid? *Brain Sci.* 2022;12(12):1729. <https://doi.org/10.3390/ijms25021168>
43. Eldem E, Barve A, Sallin O, Foucras S, Annoni JM, Schmid AW, et al. Salivary proteomics identifies transthyretin as a biomarker of early dementia conversion. *J Alzheimers Dis Rep.* 2022;6(1):31–41. <https://doi.org/10.3233/ADR-220176>
44. Marksteiner J, Defrancesco M, Humpel C. Saliva tau and phospho-tau-181 measured by Lumipulse in patients with Alzheimer's disease. *Front Aging Neurosci.* 2022; 14:1014305. <https://doi.org/10.3389/fnagi.2022.1014305>
45. Boston PF, Gopalkaje K, Manning L, Middleton L, Loxley M. Developing a simple laboratory test for Alzheimer's disease: measuring acetylcholinesterase in saliva-a pilot study. *International Journal of Geriatric Psychiatry: A journal of the psychiatry of late life and allied sciences.* 2008;23(4):439–40. <https://doi.org/10.1002/gps.1913>
46. Souza-Talarico JN de, Caramelli P, Nitrini R, Chaves EC. Effect of cortisol levels on working memory performance in elderly subjects with Alzheimer's disease. *Arq Neuropsiquiatr.* 2008; 66:619–24. <https://doi.org/10.1590/S0004-282X2008000500003>

47. Kim CB, Choi YY, Song WK, Song KB. Antibody-based magnetic nanoparticle immunoassay for quantification of Alzheimer's disease pathogenic factor. *J Biomed Opt.* 2014;19(5):51205. <https://doi.org/10.1117/1.JBO.19.5.051205>
48. Liang Q, Liu H, Zhang T, Jiang Y, Xing H, Zhang A hua. Metabolomics-based screening of salivary biomarkers for early diagnosis of Alzheimer's disease. *RSC Adv.* 2015;5(116):96074–9. <https://doi.org/10.1039/C5RA19094K>
49. Ahmadi-Motamayel F, Goodarzi MT, Tarazi S, Vahabian M. Evaluation of salivary acetylcholinesterase and pseudocholinesterase in patients with Alzheimer's disease: A case-control study. *Special Care in Dentistry.* 2019;39(1):39–44. <https://doi.org/10.1111/scd.12342>
50. Pেকেles H, Qureshi HY, Paudel HK, Schipper HM, Gornistky M, Chertkow H. Development and validation of a salivary tau biomarker in Alzheimer's disease. *Alzheimer's & Dementia: Diagnosis, Assessment & Disease Monitoring.* 2019; 11:53–60. <https://doi.org/10.1016/j.dadm.2018.03.003>
51. François M, Karpe A, Liu JW, Beale D, Hor M, Hecker J, et al. Salivaomics as a potential tool for predicting Alzheimer's disease during the early stages of neurodegeneration. *J Alzheimer's Dis.* 2021;82(3):1301–13. <https://doi.org/10.3233/JAD-201322>
52. Cui Y, Zhang H, Zhu J, Liao Z, Wang S, Liu W. Investigation of whole and glandular saliva as a biomarker for Alzheimer's disease diagnosis. *Brain Sci.* 2022;12(5):595. <https://doi.org/10.3390/brainsci12050595>
53. Ryu IS, Kim DH, Ro JY, Park BG, Kim SH, Im JY, et al. The microRNA-485-3p concentration in salivary exosome-enriched extracellular vesicles is related to amyloid  $\beta$  deposition in the brain of patients with Alzheimer's disease. *Clin Biochem.* 2023; 118:110603. <https://doi.org/10.1016/j.clinbiochem.2023.110603>
54. Contini C, Fadda L, Lai G, Masala C, Olianias A, Castagnola M, et al. A top-down proteomic approach reveals a salivary protein profile able to classify Parkinson's disease with respect to Alzheimer's disease patients and to healthy controls. *Proteomics.* 2024;24(3–4):2300202. <https://doi.org/10.1002/pmic.202300202>
55. Shi M, Sui YT, Peskind ER, Li G, Hwang H, Devic I, et al. Salivary tau species are potential biomarkers of Alzheimer's disease. *J Alzheimer's Dis.* 2011;27(2):299–305. <https://doi.org/10.3233/JAD-2011-110663>
56. Nijakowski K, Zdrojewski J, Nowak M, Gruszczynski D, Knoll F, Surdacka A. Salivary Metabolomics for Systemic Cancer Diagnosis: A Systematic Review. *Metabolites.* 2022;13(1). <https://doi.org/10.3390/metabo13010028>
57. Gouras GK, Olsson TT, Hansson O.  $\beta$ -Amyloid peptides and amyloid plaques in Alzheimer's disease. *Neurotherapeutics.* 2015;12(1):3–11. <https://doi.org/10.1007/s13311-014-0313-y>
58. Imran NK, Ali OH, Ahmed MA. Evaluation of Salivary? -Amylase Enzyme Activity in Smoker and Peptic Ulcer Patients affected by Periodontitis in Relation to Clinical Periodontal Parameters. *Indian Journal of Forensic Medicine & Toxicology.* 2020;14(4):2349–55. <https://doi.org/10.37506/ijfmt.v14i4.11939>
59. Yong SJ, Veerakumarasivam A, Lim WL, Chew J. Neuroprotective effects of lactoferrin in Alzheimer's and Parkinson's diseases: A narrative review. *ACS Chem Neurosci.* 2023;14(8):1342–55. <https://doi.org/10.1021/acchemneuro.2c00679>
60. Thapa S, Lv M, Xu H. Acetylcholinesterase: a primary target for drugs and insecticides. *Mini Rev Med Chem.* 2017;17(17):1665–76. <https://doi.org/10.2174/1389557517666170120153930>
61. Saxena M, Dubey R. Target enzyme in Alzheimer's disease: Acetylcholinesterase inhibitors. *Curr Top Med Chem.* 2019;19(4):264–75. <https://doi.org/10.2174/1568026619666190128125912>

62. Avila J, Lucas JJ, Perez MAR, Hernandez F. Role of tau protein in both physiological and pathological conditions. *Physiol Rev.* 2004. <https://doi.org/10.1152/physrev.00024.2003>
63. Peineau S, Rabiant K, Pierrefiche O, Potier B. Synaptic plasticity modulation by circulating peptides and metaplasticity: Involvement in Alzheimer's disease. *Pharmacol Res.* 2018; 130:385–401. <https://doi.org/10.1016/j.phrs.2018.01.018>
64. Heneka MT, Carson MJ, El Khoury J, Landreth GE, Brosseron F, Feinstein DL, et al. Neuroinflammation in Alzheimer's disease. *Lancet Neurol.* 2015;14(4):388–405. [https://doi.org/10.1016/S1474-4422\(15\)70016-5](https://doi.org/10.1016/S1474-4422(15)70016-5)
65. Ouanes S, Popp J. High Cortisol and the Risk of Dementia and Alzheimer's Disease: A Review of the Literature. *Front Aging Neurosci.* 2019;11:43. <https://doi.org/10.3389/fnagi.2019.00043>
66. Enad HH, Al-Mizraqchi AS. Salivary Cortisol as a Stress Biomarker and Total Viable Count of Salivary Bacterial Microbiome among COVID-19 Patients. *J Bagh Coll Dent.* 2021;33(4):6-10. <https://doi.org/10.26477/jbcd.v33i4.3013>
67. Paula-Lima AC, Brito-Moreira J, Ferreira ST. Deregulation of excitatory neurotransmission underlying synapse failure in Alzheimer's disease. *J Neurochem.* 2013;126(2):191–202. <https://doi.org/10.1111/jnc.12304>
68. Dalangin R, Kim A, Campbell RE. The role of amino acids in neurotransmission and fluorescent tools for their detection. *Int J Mol Sci.* 2020;21(17):6197. <https://doi.org/10.3390/ijms21176197>
69. Ho PTB, Clark IM, Le LTT. MicroRNA-based diagnosis and therapy. *Int J Mol Sci.* 2022;23(13):7167. <https://doi.org/10.3390/ijms23137167>
70. Carafa V, Rotili D, Forgione M, Cuomo F, Serrettiello E, Hailu GS, et al. Sirtuin functions and modulation: from chemistry to the clinic. *Clin Epigenetics.* 2016;8(1):61. <https://doi.org/10.1186/s13148-016-0224-3>
71. Pukhalskaia AE, Dyatlova AS, Linkova NS, Kozlov KL, Kvetnaia T V, Koroleva M V, et al. Sirtuins as possible predictors of aging and Alzheimer's disease development: verification in the hippocampus and saliva. *Bull Exp Biol Med.* 2020;169(6):821–4. <https://doi.org/10.1007/s10517-020-04986-4>
72. Khalifeh M, Read MI, Barreto GE, Sahebkar A. Trehalose against Alzheimer's disease: insights into a potential therapy. *BioEssays.* 2020;42(8):1900195. <https://doi.org/10.1002/bies.201900195>
73. Johnson CH, Ivanisevic J, Siuzdak G. Metabolomics: beyond biomarkers and towards mechanisms. *Nat Rev Mol Cell Biol.* 2016;17(7):451–9. <https://doi.org/10.1038/nrm.2016.25>

#### الدقة التشخيصية للمؤشرات الحيوية اللعابية في مرض الزهايمر: مراجعة منهجية عامر عبدالله مجيد ، عدي محمود الأعاني ، نذير هاشم الراوي المستخلص:

خلفه: ينسب مرض الزهايمر في التدهور المعرفي وموت الخلايا العصبية بسبب لويحات أميلويد بيتا والتشابك الليفي العصبي. طرق التشخيص الحالية تدخلية ومكلفة وقليلة ، لكن التشخيص المبكر يحسن نتائج المرضى. نظرا لعلاقة محور الفم والدماع بالجهاز العصبي المركزي ، قد يكون اللعاب مصدرا قابلا للتطبيق للمؤشرات الحيوية لمرض الزهايمر غير التداخلي. تهدف هذه المراجعة المنهجية إلى تقييم الإمكانات التشخيصية للمؤشرات الحيوية اللعابية لمرض الزهايمر من خلال مراجعة البيانات الموجودة بعناية. المنهجية : كانت الدراسات المنشورة بين 1 يناير 2008 و 30 سبتمبر 2023 موضوع بحث شامل في الأدبيات في مجلات سكوبس وشبكة العلوم وقواعد البيانات. تم استخراج البيانات ذات الصلة بعد تقييم الدراسات وفقا لمعايير التضمين والاستبعاد المحددة مسبقا. تم تنفيذ معايير مركز أكسفورد للطب القائم على الأدلة لتقييم جودة الدراسات التي تم دمجها. النتائج : تم تضمين ما مجموعه 36 دراسة في المراجعة المنهجية ، لأنها استوفت معايير الإدماج. أظهر مرضى الزهايمر مستويات لعاب مرتفعة بشكل ملحوظ من بيتا أميلويد 42 و p-Tau بالمقارنة مع الضوابط الصحية. أظهر المرضى الذين يعانون من مرض الزهايمر اتجاهات غير مهمة في اللعاب T-Tau واللاكثوفيرين. الخاتمة : تغير في مستويات بيتا أميلويد 42 و p-Tau في لعاب المرضى الذين يعانون من مرض الزهايمر بشكل كبير. تؤكد هذه النتائج على احتمال اللعاب كمصدر غير جراحي للمؤشرات الحيوية لتشخيص مرض الزهايمر. ومع ذلك ، يجب أن تخضع اختبارات اللعاب للتحقق الإضافي في مجموعات أكبر وذات خصائص جيدة وتوحيد المنهجيات قبل أن يتم تنفيذها في الممارسة السريرية.